

# Shigella

Chip-based Real Time PCR Test for Shigella spp.

#### 1. INTENDED USE

Truenat® Shigella (REF601470005 / 601470020 / 601470025 / 601470050 / 601470100 / 601470200) is a Chip-based Real Time Polymerase Chain Reaction (PCR) test for the semi-quantitative detection of *Shigella* spp. in human stool specimen and aids in the diagnosis of infection with Shigella. Truenat® Shigella runs on the Truelab® Real Time Quantitative micro PCR Analyzers. Truenat® Shigella is an *in vitro* diagnostics test meant for professional use only.

#### 2. INTRODUCTION

Shigella is a Gram-negative, facultative anaerobic, non-spore-forming, non-motile, rod-shaped bacterium which is genetically closely related to *E. coli. Shigella* causes Shigellosis in humans. During infection it typically causes dysentery. Shigella is one of the leading bacterial causes of diarrhea worldwide, causing an estimated 80-165 million cases. The number of deaths it causes each year is estimated at between 74,000 and 600,000. It is one of the top four pathogens that cause moderate-to-severe diarrhea in African and South Asian children. Three Shigella groups are the major disease-causing species, among them S. flexneri is the most frequently isolated species worldwide, and accounts for 60% of cases in the developing world; S. sonnei causes 77% of cases in the developed world, compared to only 15% of cases in the developing world, and S. dysenteriae is usually the cause of epidemics of dysentery, particularly in confined populations such as refugee camps.

Shigella species generally invade the epithelial lining of the colon, causing severe inflammation and death of the cells lining the colon. This inflammation results in diarrhea and even dysentery that are the hallmarks of Shigella infection. Some strains of Shigella produce toxins which contribute to the disease during infection. S.flexneri strains produce ShET1 and ShET2 toxins, which contribute to diarrhea. S.dysenteriae strains produce Shiga toxin, which is hemolytic similar to the verotoxin produced by enterohemorrhagic E. coli. Both Shiga toxin and verotoxin are associated with causing potentially fatal hemolytic-uremic syndrome. The most common symptoms are diarrhea, fever, nausea, vomiting, stomach cramps, and flatulence. It is also commonly known to cause large and painful bowel movements.

The stool may contain blood, mucus or pus. Hence, *Shigella* cells may cause dysentery. In rare cases, young children may have seizures. Symptoms can take as long as a week to appear, but most often begin two to four days after ingestion. Symptoms usually last for several days, but can last for weeks. *Shigella* is implicated as one of the pathogenic causes of reactive arthritis worldwide.



Early and accurate diagnosis of shigellosis coupled with prompt medical intervention is essential for reducing the morbidity and mortality caused by *Shigella* spp. Laboratory detection of *Shigella* species involves isolation on selective media, followed by biochemical tests, such as indole and lysine decarboxylase to differentiate *Shigella*. Species identification is further confirmed by serotyping of the surface antigens. The Current laboratory diagnosis of *Shigella* spp. is laborious and time consuming and has low sensitivity. With the advent of molecular testing, accurate detection of *Shigella* has increased. However, molecular tests such as PCR have so far been restricted to centralized reference laboratories as they require skilled manpower and elaborate infrastructure. Also the turnaround time for results could take a few days to weeks leading to high losses to follow-up.

The **Truelab**® Real Time Quantitative micro PCR System enables decentralization and near patient detection and diagnosis of Shigella by making real time PCR technology rapid, simple, robust and user friendly and offering "sample to result" capability even at resource limited settings. This is achieved through a combination of lightweight, portable, mains / battery operated **Truelab**® Real Time micro PCR Analyzer and **Trueprep® AUTO/AUTO v2** Universal Cartridge Based Sample Prep Device and room temperature stable **Truenat®** micro PCR chips and **Trueprep® AUTO/AUTO v2** Universal Cartridge Based Sample Prep Kits so that even the peripheral laboratories with minimal infrastructure and minimally trained technician can easily perform these tests routinely in their facilities and report PCR results in less than an hour. Moreover, with these devices PCR testing can also be initiated in the field level, on site.

**Truenat® Shigella** is a disposable, room temperature stable, micro PCR test with dried MgCl₂ in reaction well and freeze dried PCR reagents in microtube for performing Real Time PCR test for Shigella virus and runs on the **Truelab®** Real Time Quantitative micro PCR Analyzer. It requires only six (6) µL of purified DNA to be added to the reaction well for the analysis. The intelligent chip also carries test and batch related information. The **Truenat® Shigella** chip also stores information of used chips to prevent any accidental re-use of the chip.

NOTE: Truelab® / Truenat® / Trueprep® / Truepet® are all trademarks of Molbio Diagnostics Private Limited.

The Truelab® Real Time micro PCR Analyzer is protected by the following

patents and patents granted: IN 2313/CHE/2007 (Patent No. 281573), WO2009/047804 and corresponding claims of any foreign counterpart(s) thereof.

The Truenat<sup>®</sup> micro PCR chip is protected by the following patents and patents pending: IN 2312/CHE/2007, WO 2009/047805 and corresponding claims of any foreign counterpart(s) thereof.

### 3. PRINCIPLE OF THE TEST

Truenat® Shigella works on the principle of Real Time Polymerase Chain Reaction based on Tagman chemistry. The DNA from the patient sample is first extracted using Trueprep® AUTO/AUTO v2 Universal Cartridge Based Sample Prep Device and Trueprep® AUTO/AUTO v2 Universal Cartridge Based Sample Prep Kit. The Truenat Shigella chip is placed on the chip tray of the Truelab Real Time Quantitative micro PCR Analyzer. Six (6) µL of the purified DNA is then dispensed using the provided micropipette and tip into the microtube containing freeze dried PCR reagents and allowed to stand for 30-60 seconds to get a clear solution. Δ No mixing by tapping, shaking or by reverse pipetting should be done. Six (6) μL of this clear solution is then pipetted out using the same pipette and tip and dispensed into the reaction well of the Truenat® Shigella chip and the test is started. A positive amplification causes the dual labeled fluorescent probe in the **Truenat® Shigella** chip to release the fluorophores in an exponential manner which is then captured by the built-in opto-electronic sensor and displayed as amplification curve on the analyzer screen, on a real time basis during the test run. The Cycle threshold (Ct) is defined as the number of amplification cycles required for the fluorescent signal to cross the threshold (i.e. exceed the background signal). Ct levels are inversely proportional to the amount of target nucleic acid in the sample (i.e. the lower the Ct level the greater is the amount of target nucleic acid in the sample). In the case of negative samples, amplification does not occur and a horizontal amplification curve is displayed on the screen during the test run. At the end of the test run, Shigella "DETECTED" or "NOT DETECTED" result is displayed and in positive cases, semi-quantitative result is also displayed on the screen. Based on the detection of the internal positive control (IPC), the validity of the test run is also displayed. The IPC is a full process control that undergoes all the processes the specimen undergoes - from extraction to amplification thereby validating the test run from sample to result. Absence of or shift of IPC Ct beyond a pre-set range in case of negative samples invalidates the test run. While IPC will co-amplify in most positive cases also, in some specimen having a high target load, the IPC may not amplify, however the test run is still considered valid. The results can be printed using the Truelab® micro PCR printer or transferred to the lab computer/or any remote computer via Wifi network or 3G/GPRS network. Upto 20000 results in Truelab® Uno Dx / Duo / Quattro can be stored on the analyzer for future recall and reference.

## 4. TARGET SELECTION

The gene target sequences for this assay is *ipgF* gene producing invasion associated protein of Shigella genome.

#### 5. CONTENTS OF THE Truenat® Shigella KIT

- A. Individually sealed pouches, each containing
  - 1. Truenat® Shigella micro PCR chip.
  - 2. Microtube with freeze dried PCR reagents.
  - 3. DNase & RNase free pipette tip.
  - Desiccant pouch
- B. Package Insert.

REF	601470005	601470020	601470025	601470050	601470100	601470200
Σ	5T	20T	25T	50T	100T	200T

## 6. CONTENTS OF THE Trueprep® AUTO Universal Sample Pre-treatment Pack

- A. Lysis buffer.
- B. Disposable transfer pipette (graduated).
- C. Package Insert.

REF	60205AB05	60205AB20	60205AB25	60205AB50	60205AB100	60205AB200
\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\\	5T	20T	25T	50T	100T	200T

#### 7. STORAGE AND STABILITY

**Truenat® Shigella** chip is stable for two (2) years from the date of manufacture if stored between 2-30°C. It is also stable for one (1) month at temperatures up to 45°C. Avoid exposure to light or elevated temperatures (above recommended levels). Do not freeze.

**Trueprep® AUTO** Universal Sample Pre-treatment Pack is stable for two (2) years from the date of manufacture if stored between 2-40°C. It is also stable for one (1) month at temperatures upto 45°C. Do not freeze.

#### 8. MATERIALS REQUIRED BUT NOT PROVIDED WITH THE KIT

**Truelab**® Real Time micro PCR Workstation (REF 623010001 / 633010001 / 643010001/653010001) consisting of

1. Trueprep® AUTO/AUTO v2 Universal Cartridge Based Sample Prep Device

- (REF603041001/603042001)
- Truelab<sup>®</sup> Uno Dx / Truelab<sup>®</sup> Duo / Truelab<sup>®</sup> Quattro Real Time micro PCR Analyzer (REF 603021001/603022001/603023001).
- Truelab<sup>®</sup> micro PCR Printer (REF 603050001).
- Truepet® SPA fixed volume precision micropipette 6 µl (REF 604070006).
- 5. Truelab® Microtube Stand (REF 603070001).

Also required additionally are: Trueprep® AUTO Universal Sample Pre-treatment Pack (REF60205AB05 / REF60205AB20 / REF60205AB25 / REF60205AB50 / REF60205AB100 / REF60205AB200), Trueprep® AUTO Universal Cartridge Based Sample Prep Kit (REF60203AR05 / REF60203AR25 / REF60203AR50 / REF60203AR100) or Trueprep® AUTO v2 Universal Cartridge Based Sample Prep Kit (REF60207AR05 / REF60207AR25 / REF60207AR50 / REF60207AR100), Truenat® Positive Control Kit - Panel V (REF 801050008), Nylon flocked swabs, Powder free disposable gloves, waste disposal container with lid.

### SPECIMEN PREPARATION FOR EXTRACTION WITH Trueprep® AUTO/AUTO v2 Stool specimen:

**For Solid stool specimens:** Transfer solid stool sample (approximately 100-150 mg) using an appropriate disposable swab / spatula / wooden applicator stick into the lysis buffer tube. Dispose of the used swab / spatula / wooden applicator stick as per the section on "Disposal and Destruction" (Section 17). Mix the contents of the lysis buffer by vortexing for 1 minute. Allow the contents of the tube to settle at room temperature for 5 minutes.

For Watery stool specimens: Transfer 150 µL of the watery stool specimen using a suitable micropipette/Pasteur pipette into the lysis buffer tube. Dispose of the used pipette tip/Pasteur pipette as per the section on "Disposal and Destruction" (Section 17). Mix the contents of the lysis buffer by vortexing for 1 minute. Allow the contents of the tube to settle at room temperature for 5 minutes. ⚠ Dispose off the used swab / spatula / Wooden applicator stick / pipette tip / Pasteur pipette as per the section on "Disposal and Destruction" (Section 17). (Refer to package insert of **Trueprep**® **AUTO** Universal Sample Pre-treatment Pack for details).

#### Sample Storage and Transportation:

Sample Pre-treatment decontaminates the specimen and makes it ready for storage/ transportation/extraction. The specimen in this form is stable for up to three (3) days at 40°C and one (1) week at 30°C.

Nucleic acid extraction: Transfer (1.5 to 2 ml) of the clear suspension to cartridge using the transfer pipettes provided with Trueprep® AUTO/AUTO v2 Universal Cartridge Based Sample Prep Kit. Ensure that no particulate matter is transferred from the suspension in the lysis buffer to the cartridge. Dispose of lysis buffer tube and transfer pipette after use, as per the section on "Disposal and Destruction" (Section 17). Carry out further extraction procedure with the Trueprep® AUTO/AUTO v2 Universal Cartridge Based Sample Prep Device. (Refer to the User Manual of Trueprep® AUTO/AUTO v2 Universal Cartridge Based Sample Prep Device and the package insert of Trueprep® AUTO/AUTO v2 Universal Cartridge Based Sample Prep Kit for details).

## 10. SAFETY PRECAUTIONS

- 1. For in vitro diagnostic use only.
- 2. Bring all reagents and specimen to room temperature (20 30°C) before use.
- 3. Do not use kit beyond expiry date.
- Carefully read the User Manuals, package inserts and Material Safety Data Sheets (MSDS) of all the components of the Truelab<sup>®</sup> Real Time micro PCR System before use.
- 5. All materials of human origin should be handled as though potentially infectious.
- 6. Do not pipette any material by mouth.
- Do not eat, drink, smoke, apply cosmetics or handle contact lenses in the area where testing is done.
- Use protective clothing and wear disposable gloves when handling samples and while performing sample extraction.

## 11. PROCEDURAL PRECAUTIONS

- Check all packages before using the kit. Damage to the packaging does not prevent the contents of the kit from being used. However, if the outer packaging is damaged the user must confirm that individual components of the kit are intact before using them.
- Do not perform the test in the presence of reactive vapours (e.g. from sodium hypochlorite, acids, alkalis or aldehydes) or dust.
- While retrieving the Truenat<sup>®</sup> Shigella micro PCR chip, microtube and the DNase and RNase free pipette tip from the pouch, ensure that neither bare hands nor gloves that have been used for previous tests run are used.

### 12. PROCEDURAL LIMITATIONS

- Optimal performance of this test requires appropriate specimen collection, handling, storage and transport to the test site.
- 2. Though very rare, mutations within the highly conserved regions of the target genome where the **Truenat**® assay primers and/or probe bind may result in the under-quantitation of or a failure to detect the presence of the concerned pathogen.
- 3. The instruments and assay procedures are designed to minimize the risk of

- contamination by PCR amplification products. However, it is essential to follow good laboratory practices and ensure careful adherence to the procedures specified in this package insert for avoiding nucleic acid contamination from previous amplifications, positive controls or specimens.
- 4. A specimen for which the **Truenat**<sup>®</sup> assay reports "Not Detected" cannot be concluded to be negative for the concerned pathogen. As with any diagnostic test, results from the **Truenat**<sup>®</sup> assay should be interpreted in the context of other clinical and laboratory findings.

### 13. CLEANING AND DECONTAMINATION

- Spills of potentially infectious material should be cleaned up immediately with absorbent paper tissue and the contaminated area should be decontaminated with disinfectants such as 0.5% freshly prepared sodium hypochlorite [10 times dilution of 5% sodium hypochlorite (household bleach)] before continuing work.
- Sodium hypochlorite should not be used on an acid-containing spill unless the spill-area is wiped dry first. Materials used to clean spills, including gloves, should be disposed off as potentially bio-hazardous waste e.g. in a biohazard waste container.

#### 14. TEST PROCEDURE

(Please also refer the **Truelab**® Real Time Quantitative micro PCR Analyzer user manual).

- 1. Switch on the **Truelab**<sup>®</sup> Analyzer.
- 2. Select user and enter password.
- 3. For **Truelab**® **Uno Dx**, select the test profile for "Shigella" to be run from the Profiles Screen on the Analyzer screen. For **Truelab**® **Duo/Quattro**, select the Bay (Idle1/2) for **Duo** and (Idle1/2/3/4) for **Quattro** from the Status Screen to view the Profiles Screen. Select the test profile for "Shigella" to be run from the Profiles Screen on the Analyzer screen.
- 4. Enter the patient details as prompted in the **Truelab®** Analyzer screen.
- 5. Press Start Reaction.
- For Truelab<sup>®</sup> Uno Dx, Press the eject button to open the chip tray. For Truelab<sup>®</sup> Duo/Quattro, the chip tray opens automatically on tapping the "Start Reaction" button.
- Open a pouch of Truenat® Shigella and retrieve the micro PCR chip, microtube and DNase & RNase free pipette tip.
- Place the Truenat<sup>®</sup> Shigella chip on the chip tray without touching the white reaction well. The reaction well should be facing up and away from the Analyzer. Gently press the chip to ensure that it has seated in the chip tray properly.
- 9. Place the microtube containing freeze dried PCR reagents in the microtube stand provided along with the Truelab® Real Time micro PCR workstation after ensuring that white pellet of dried PCR reagents remains at the bottom of the microtube. Remove the microtube cap and dispose it off as per the section on "Disposal and Destruction" (Section 17). Using the filter barrier tip provided in the pouch, pipette out six (6) μL of the purified DNA from the Elute Collection Tube into the microtube. Allow it to stand for 30-60 seconds to get a clear solution. Δ Do not mix it by tapping, shaking or by reverse pipetting. Using the same filter barrier tip, pipette out six (6) μL of this clear solution and dispense into the centre of the white reaction well of the Truenat® Shigella chip. Take care not to scratch the internal well surface and not to spill elute on the outside of the well. Dispose off the microtip as per the section on "Disposal and Destruction" (Section 17).
- 10. For Truelab® Uno Dx, slide the chip tray containing the Truenat® Shigella Chipbased Real Time PCR test loaded with the sample into the Truelab® Analyzer. Press Done on the "Please Load Sample" Alert message. For Truelab® Duo/Quattro, select "YES" at the Please load Sample prompt. Chip tray will close automatically and the reaction will start.
- 11. Read the result from the screen.
- After the reaction is completed, for Truelab<sup>®</sup> Uno Dx, push the Eject button to eject the chip tray. For Truelab<sup>®</sup> Duo/Quattro, tap the "Open/Close Tray" button to eject the chip tray.
- Take out the Truenat<sup>®</sup> Shigella Chip-based Real Time PCR test at end of the test and dispose it off as per the section on "Disposal and Destruction" (Section 17).
- 14. Turn on Truelab® micro PCR printer and select print on the screen for printing out hard copy of the results. Test results are automatically stored and can be retrieved any time later. (Refer to the Truelab® Analyzer manual).
- 15. Switch off the **Truelab**® Analyzer.

## 15. RESULTS & INTERPRETATIONS

Two amplification curves are displayed on the **Truelab**® Analyzer screen to indicate the progress of the test. Both the target and the internal positive control (IPC) curves will take a steep, exponential path when the fluorescence crosses the threshold value in case of positive samples. The Cycle threshold (Ct) will depend on the number of target nucleic acids in the sample. The curve will remain horizontal throughout the test duration and the IPC curve will take an exponential path in case of negative samples. In case the IPC curve remains horizontal in a negative sample, the test is considered as Invalid. At the end of the test run, the results screen will display "DETECTED" for Positive result or "NOT DETECTED" for Negative result. The result

screen would also display the microbial load as "HIGH (Ct<20)", "MEDIUM ( $20 \le Ct < 25$ )", "LOW ( $25 \le Ct < 30$ )" or "VERY LOW (Ct  $\ge 30$ )" for positive specimen. The result screen also displays the validity of the test run as "VALID" or "INVALID". Invalid samples have to be repeated with fresh specimen from the sample preparation stage. \*While IPC will co-amplify in most positive cases also, in some specimen having a high target load, the IPC may not amplify, however the test run is still considered valid.

#### 16. QUALITY CONTROL PROCEDURES

To ensure that the **Truelab**® Real Time Quantitative micro PCR Analyzer is working accurately, run positive and negative controls from time to time. **Truenat**® Positive Control Kit - Panel V (REF 801050008) containing Positive Control and Negative Control must be ordered separately. It is advisable to run controls under the following circumstances. ◆ Whenever a new shipment of test kits is received. ◆ When opening a new test kit lot. ◆ If the temperature of the storage area falls outside of 2-30°C. ◆ By each new user prior to performing testing on clinical specimen.

#### 17. DISPOSAL AND DESTRUCTION

- Submerge the used Truenat<sup>®</sup> Shigella chip, microtube, microtube cap, transfer pipette, pipette tips, lysis buffer tube, nylon flock swab etc. in freshly prepared 0.5 % sodium hypochlorite solution for 30 minutes before disposal as per the standard medical waste disposal guidelines.
- 2. Disinfect the solutions and/or solid waste containing biological samples before discarding them according to local regulations.
- 3. Samples and reagents of human and animal origin, as well as contaminated materials, disposables, neutralized acids and other waste materials must be discarded according to local regulations after decontamination by immersion in a freshly prepared 0.5% of sodium hypochlorite for 30 minutes (1 volume of 5% sodium hypochlorite for 10 volumes of water).
- 4. Do not autoclave materials or solutions containing sodium hypochlorite.
- Chemicals should be handled in accordance with Good Laboratory Practice and disposed off according to the local regulations.

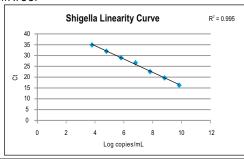
#### 18. SPECIFIC PERFORMANCE CHARACTERISTICS

Analytical Exclusitivity (Primer specificity): The following microorganisms were evaluated in *silico* from the NCBI database using the NCBI nucleotide blast and primer blast tools to determine for potential cross-reactivity in the Truenat® Shigella assay. Results obtained showed no cross reactivity of the Truenat® Shigella test with the listed organisms.

Organism	Organism
Acinetobacter anitratus	Adenovirus
Enterobacter cloacae	Hepatitis C virus
Chlamydia trachomatis	Human Immunodeficiency virus
Candida albicans	Hepatitis B virus
Escherichia coli	Herpes Simplex virus
Salmonella enterica	Epstein-Barr virus
Streptococcus mutans	Simian virus
Staphylococcus aureus	Cytomegalovirus
Enterococcus faecalis	Human Papilloma virus
Trichomonas vaginalis	
Gardnerella vaginalis	
Yersenia enterocolitica	
Mycobacterium tuberculosis	
Klebsiella pneumoniae	
Pseudomonas trivialis	
Neisseria gonorrhoeae	
Mycobacterium bovis	

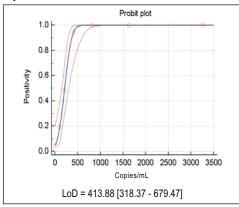
#### Linearity:

The linearity analysis was performed according to CLSI Guidelines. Serial dilutions of DNA from *Shigella sonnei* (Levine) Weldin (ATCC° 11060™) culture from ATCC ranging from 6.52E+09 to 6.52E+03 copies/mL was made in stool samples and nucleic acids were extracted on **Trueprep® AUTO** Universal Cartridge Based Sample Prep Device in triplicates for each dilution followed by PCR on **Truelab®** Real Time micro PCR analyzer using **Truenat® Shigella** test. The assay is found to be linear over 7 orders of magnitude (from 6.52E+09 to 6.52E+03 copies/mL) for **Truenat® Shigella** DNA from *Shigella sonnei* (Levine) Weldin (ATCC® 11060™) culture from ATCC.



#### Limit of detection (LoD):

The LoD was determined by spiking dilutions of *Shigella sonnei* (Levine) Weldin (ATCC° 11060™) culture DNA from ATCC into stool samples and performing on **Trueprep® AUTO** Universal Cartridge Based Sample Prep Device followed by PCR on **Truelab®** Real Time micro PCR analyzer using **Truenat® Shigella** test. Probit analysis of the data was used to determine the concentration of the DNA with 95% probability of detection. The LoD was found to be 413.88 copies/mL for **Truenat® Shigella** assay.



#### Robustness:

Potential sample carryover within the **Truenat**® **Shigella** test was evaluated by testing alternate positive followed by negative samples. The numbers of samples runs were 20 positives and 20 negatives. The results showed no carryover contamination. The **Truenat**® **Shigella** test did not exhibit any detectable carry over contamination between positive and negative samples.

## Reproducibility:

The purpose of this study is to compare the functional performance of the **Truenat**<sup>®</sup> **Shigella** assay using three different titres of samples on **Truelab**<sup>®</sup> Real Time micro PCR analyzer. High, Medium and low titre samples were extracted on **Trueprep**<sup>®</sup> **AUTO** Universal Cartridge Based Sample Prep Device and tested among three different users (Inter user), on three different devices (Inter device) and on 5 consecutive days (Inter day) to check the variability. Mean %CV values for all titres has been calculated for Inter User (1.79), Inter day (1.79) and Inter Device (1.98) which were in the accepted range of ≤15% CV for **Truenat**® **Shigella** assay.

#### Interference

The purpose of this study is to determine the effect of interfering substances on the **Truenat** Shigella assay. The experiments were performed with Shigella culture DNA spiked into stool samples. Potentially interfering substances used in this study are Albumin: 9 g/dL, Triglycerides: 3.0 mg/dL, Human DNA: 0.4 mg/dL, Hemoglobin: 500 mg/dL. The presence of potentially interfering substances did not interfere with the performance of **Truenat** Shigella assay. The CV values obtained were within the accepted range of ≤15% for **Truenat** Shigella assay.

## Precision:

Precision was tested by performing **Truenat** Shigella assay with extracted DNA of High, Medium and Low titres for five consecutive days. Every day PCR for each titre DNA was run in duplicates. The %CV values obtained for High titre (5.32), Medium titre (3.80) and low titre (3.32) were within the accepted range of ≤15% CV for **Truenat** Shigella assay.

### Clinical validation:

A panel of 30 stool samples comprising of 20 negative and 10 positive specimens were tested on three different manufacturing lots of **Truenat® Shigella** assay at Ramaiah Medical College Hospital Laboratory, Bangalore against the SACACE Shigella/Salmonella/Campylobacter Real-TM PCR kit as the reference test.

	SACACE Shigella/Salmonella/Campylobacter Real-TM PCR Kit					
		Positive	Negative	Total		
Truenat <sup>®</sup> Shigella	Positive	10	0	10		
Tructiut Singena	Negative	0	20	20		
	Total	10	20	30		

With the consideration of above data, **Truenat**® **Shigella** test performed consistently in this study with observed sensitivity of 100% and specificity of 100% in comparison with SACACE Shigella/Salmonella/Campylobacter Real-TM PCR reference kit and the inter lot variation data obtained was within the accepted range of ≤15% CV for the **Truenat**® **Shigella** test.

## 19. REFERENCES

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